

S-Trap™ HYPERsol FFPE with PIXUL Megasonication Protocol

Tissue	Expected µg Soluble Protein/mg
Lymph Node	117
Heart	111
Pancreas	114
Colon	97
Spleen	96
Thyroid	93
Liver	84
Parathyroid	77
Cerebellum	77

Tissue	Expected µg Soluble Protein/mg
Cerebral Cortex	81
Kidney	74
Stomach	71
Uterus	60
Marrow	49
Testis	47
Prostate	43
Adrenal Gland	42
Lung	39

Protocol:

- Remove Excess Paraffin.** Trim FFPE blocks of excess paraffin using a sterile razor blade or scapula.
- Punch or Section.** Use a Biopsy Punch (for example a 1 mm Kai Miltex-Integra 33-31AA) to punch cores or alternatively section scrolls on a microtome. 5 mg of total FFPE tissue is recommended; pool as necessary.
- Solubilize and Decrosslink.**
 - For tissue cores, dice them into small pieces with a scapula or razor blade then transfer to a 96-well plate. For scrolls, transfer to a 96-well plate; use tweezers to position on the bottom of the well.
 - In the EU and Japan, completely remove paraffin from the sample with an organic solvent such as xylene or methyl tert-butyl ether (MTBE): add 100 µL, shake samples for 5 minutes at room temperature then remove all organic solvent to yield deparaffinized samples. Repeat as necessary to completely remove paraffin.
 - Add 100 µL of S-Trap™ Solution 1, 2x Lysis Solution to 5 mg FFPE tissue or, if starting with less, add 20.0 µL per 1 mg.
 - Allow samples to rehydrate overnight.
 - Mega-sonicate at room temperature for 5 minutes on the PIXUL or until pieces have been dissolved: 50 N pulse at 1 kHz with a 20 Hz burst rate.
 - Place on a heat block at 80 °C for 1 hour to reverse crosslinking. Make sure to poke small holes in the plate seal to relieve pressure and spin down condensate.
 - Mega-sonicate for 6 minutes with the same settings as outlined above.
- Match Protein Loading.** Measure protein concentration with a BCA assay and match the concentration of protein in all samples by dilution as needed with S-Trap™ Solution 1, 2x Lysis Solution.
- Process per standard S-Trap™ protocol.