

S-Trap™ 96-well Mini Plate



The Protifi™ S-Trap™ technology combines robust Sodium Dodecyl Sulfate (SDS) solubilization of biological samples and enhanced contaminant removal for proteomic analysis by mass spectrometry. **S-Trap™ 96-well Mini Plates** allow for the solubilization, reduction, alkylation, and digestion of **100 µg – 300 µg of total protein** cleaned and ready for peptide analysis.

Additional Reagents/Solutions (Not Included)

Liquid Chromatography-Mass Spectrometry (LC-MS) grade is recommended, where applicable

- Protein sample (**100 µg - 300 µg of total protein**)
- Protease of choice (e.g. Trypsin or Trypsin/Lys-C)*
- LC-MS grade Water
- LC-MS grade Isopropyl Alcohol (IPA)
- LC-MS grade Methanol (MeOH)
- LC-MS grade Acetonitrile (ACN)
- LC-MS grade Formic Acid (FA)
- SDS
- 1 M Triethylammonium Bicarbonate (TEAB), pH 8.5
- (Tris(2-carboxyethyl)phosphine) (TCEP)
- Methyl Methanethiosulfonate (MMTS)
- Phosphoric Acid
- Benzonase® (optional)*

* Benzonase® and proteases are susceptible to freeze-thaw cycles.

Equipment/Materials (Not Included)

- pH meter with reference solutions
- Balance
- Single- and/or multi-channel pipettes (P2 through P1000) and corresponding tips
- Centrifuge capable of processing 96-well plates at 2000 x g
- Vortex mixer
- Heat block (adjusted to 55 °C)
- pH paper (pH < 1, optional)
- Humidified incubator (adjusted to 47 °C)
- Lyophilizer or SpeedVac
- Positive pressure apparatus or vacuum manifold (optional)
- Sonicator (optional, recommended)
- 1.0 mL and 2.0 mL square well 96-well plates
- 96-well cover

Stock Solutions and Reagent to Prepare PRIOR to Start:

All indicated solutions (**) can be purchased at protifi.com

Protein Sample	Your sample	100 µg – 300 µg
Protease	Your choice of protease (E.g., Trypsin or Trypsin/Lys-C)	1 µg - 10 µg per 100 µg of sample in Solution 6 (Digestion Solution)
Benzonase (optional)	Benzonase® and MgCl ₂	Benzonase & 50 mM MgCl ₂
Solution 1**	2x Lysis Solution	10% (w/v) SDS, 100 mM TEAB in LC-MS grade Water, pH 7.55
Solution 2**	Reductant	120 mM TCEP in LC-MS grade Water
Solution 3**	Alkylator	500 mM MMTS in LC-MS grade IPA (mass shift is + 45.988, addition of SCH ₂)
Solution 4**	Acidifier	27.5% (v/v) Phosphoric Acid in LC-MS grade Water*
Solution 5**	Binding/Wash Solution	100 mM TEAB (final) in 90% LC-MS grade MeOH, pH 7.5
Solution 6**	Digestion Solution	50 mM TEAB in LC-MS grade Water, pH 8.0
Solution 7 (optional)**	Elution Solution	0.2% (v/v) LC-MS grade FA in LC-MS grade Water
Solution 8 (optional)	Elution Solution 2	50% (v/v) LC-MS grade ACN in 0.2% (v/v) LC-MS grade FA

All solutions can be made in advance and stored at -20 °C or colder. After thawing, mix thoroughly to ensure any precipitants are redissolved.

* Acidifier must be stored **tightly capped**; it will degrade upon atmospheric exposure.

RT = room temperature (20-25 °C), v/v = volume-to-volume ratio, w/v = weight-to-volume ratio, w/w = weight-to-weight ratio

Protocol:

Refer to Appendices for Sample-Specific Considerations, Alternative Proteases, and Troubleshooting Tips.

1. **Denature Protein.** To 23.0 μ L of liquid sample, add 23.0 μ L of **Solution 1 (2x Lysis Solution)**. For solid samples, dilute **Solution 1 (2x Lysis Solution)** to 1x by adding an equal volume of LC-MS grade water. Add 46.0 μ L of 1x Solution 1 to a solid sample. Sonicate or vortex to fully solubilize and denature proteins. Samples can be processed in tubes or a deep well 96-well plate.
2. If sample is viscous due to the presence of DNA, sheer it thoroughly by probe sonication or enzymatically with a nuclease such as Benzonase™ (see Appendix D). Unsheared DNA will clog the protein trap.
3. Clarify sample as desired by centrifugation (e.g. 13,000 x g for 8 minutes). Transfer clarified lysate to tubes or a deep well 96-well plate. Pellet can be analyzed separately (see Appendix D).
4. **Reduce.** Add 2.0 μ L of **Solution 2 (Reducant)**. Incubate at 55 °C for 15 minutes.
5. **Alkylate.** Add 2.0 μ L of **Solution 3 (Alkylator)**. Incubate at RT for 10 minutes.
6. **Acidify.** Add 5.0 μ L of **Solution 4 (Acidifier)** to the sample. pH paper can be used to ensure pH ≤ 1. Proceed to the next step immediately.
7. Add 350 μ L of **Solution 5 (Binding/Wash Solution)** to the sample. Mix thoroughly.

Note: The sample may appear translucent at this step due to colloidal protein formation. Do NOT centrifuge.
8. Place an **S-Trap™ 96-well Mini Plate** atop a 2.0 mL 96-well plate. Transfer each sample, including any insoluble material, into a well of the **S-Trap™ 96-well Mini Plate**.

Note: No pre-equilibration is necessary. Solution may begin to drip through immediately; this is expected. The well can hold ~450 μ L of solution. For larger volumes, the well can be loaded multiple times with the solution from Step 7. When transferring into the well, do not disturb the matrix.
9. **Trap Protein.** Centrifuge the **S-Trap™ 96-well Mini Plate** and waste plate at 2,000 x g for 2 minutes. Visually confirm that all solution passed through the well. If not, centrifuge again for 5 minutes until no liquid remains. Discard flow-through and return the plate to the 2.0 mL 96-well plate.

Note: A vacuum manifold or positive pressure can be used to draw solution through the plate provided that all wells have flow behavior, which might not be the case for all samples.
10. **Clean Protein.** Add 250 μ L of **Solution 5 (Binding/Wash Solution)** to the **S-Trap™ 96-well Mini Plate** and centrifuge at 2,000 x g for 2 minutes. Visually confirm that all solution passed through the well. Repeat washes 3 times; discard flow-through as necessary. Return the plate to the 2.0 mL 96-well plate.

Note: Additional wash(es) may be performed if desired. Washes may be captured with the flow-through. A vacuum manifold or positive pressure may also be used if the wells have similar flow behavior.
11. Move the **S-Trap™ 96-well Mini Plate** atop a 1.0 mL 96-well plate for collection.
12. Dilute trypsin or trypsin/Lys-C in **Solution 6 (Digestion Solution)** to a final volume of 125 μ L, ensuring that the amount of protease in this solution is at a 1:10 – 1:100 (w/w) ratio with the total amount of protein in the sample (e.g. 1 μ g - 10 μ g per 100 μ g of sample).
13. Transfer the entire 125 μ L of trypsin or trypsin/Lys-C solution to the **S-Trap™ 96-well Mini Plate**, ensuring the matrix is fully covered by digestion solution.
14. **Incubate & Digest.** Loosely cover the **S-Trap™ 96-well Mini Plate** to limit evaporative loss without making an air-tight seal. Place the plate atop the 1.0 mL 96-well plate in a 47 °C humidified incubator for 1 hour. After 1 hour, add 80.0 μ L of **Solution 6 (Digestion Solution)** to all wells. Loosely cover and place back in a 47 °C humidified incubator for 1 hour.

Note: Some dripping may occur during incubation; this is not of concern. Do NOT shake.
15. **Elute 1.** Add 80.0 μ L of **Solution 6 (Digestion Solution)** to all wells of the **S-Trap™ 96-well Mini Plate** atop the 1.0 mL 96-well plate after incubation. Centrifuge at 2,000 x g for 2 minutes or until all solution has passed through.
16. **Elute 2.** Add 80.0 μ L of **Solution 7 (Elution Solution)** to all wells of the **S-Trap™ 96-well Mini Plate**. Centrifuge at 2,000 x g for 2 minutes.
17. **Elute 3.** If hydrophobic peptides are of interest, add 80.0 μ L of **Solution 8 (Elution Solution 2)** to all wells of the **S-Trap™ 96-well Mini Plate**. Centrifuge at 2,000 x g for 2 minutes.
18. Lyophilize or SpeedVac eluted peptides collected from Steps 15-17 in the 1.0 mL 96-well plate. Resuspend as needed for subsequent analysis (e.g. Aqueous Buffer A such as 5% ACN, 0.1% FA).